

High-Throughput Analysis of Opiates in Hair Samples using Laser Diode Thermal Desorption (LDTD®) coupled to Mass Spectrometry

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OVERVIEW

Ultra-Fast analysis of Opiates in hair samples using LDTD®-MS/MS

Method

- Standards, QC and samples preparation using hair matrix
- Solid-Phase Extraction (SPE)
- Deposit of a small volume of the organic phase in LazWell™ plate
- Fast Analysis using LDTD®-MS/MS system
- Cross validation LC vs LDTD®

Results

- Excellent linearity over the calibration range (R² ≥ 0.9967)
- Accuracy ranging from 93,1 to111.1%
- Precision ranging from 0.7 to 13.8 %
- Good sample stability (Wet and Dry in LazWell™)
- All samples are analyzed with a run time of 9 seconds using LDTD®-MS/MS system.

INTRODUCTION

Since the hair root is vascularized during its growth, illicit drugs present in the blood stream may enter the hair shaft via the root where they will be sequestered. Therefore, the use of illicit drugs can be revealed by analyzing a small hair sample. To increase the analysis throughput of hair samples, the laser diode thermal desorption (LDTD®) coupled to tandem mass spectrometry (MS/MS) was used for the identification and quantification of opiates.

The detection and quantification of drugs in hair samples is traditionally performed by LC-MS/MS or GC-MS analysis that require several minutes due to separation time. The LDTD®-MS/MS instrument significantly reduces analysis time and this increases the sample throughput with runtimes of 9 seconds sample-to-sample. In this study, the goal was to validate a quantitative method for the following opioid drugs in hair: 6acetylmorphine (6-AM), codeine (COD), morphine (MOR), hydrocodone (HYC), hydromorphone (HYM), oxycodone OXC) and oxymorphone (OXM), using the LDTD® coupled to MS/MS.

LDTD® Ionization Source:

The LDTD® uses a Laser Diode to produce and control heat on the sample support (Figure 1) which is a 96-well plate. The energy is then transferred through the sample holder. The sample gets dried and vaporized prior being carried by a gas in a corona discharge region. This type of ionization is characterized by a strong resistance to ionic suppression because of the absence of solvent. LDTD® ionization reduces sample-to-sample analysis time to 9 seconds and allows high throughput capabilities without carry over.

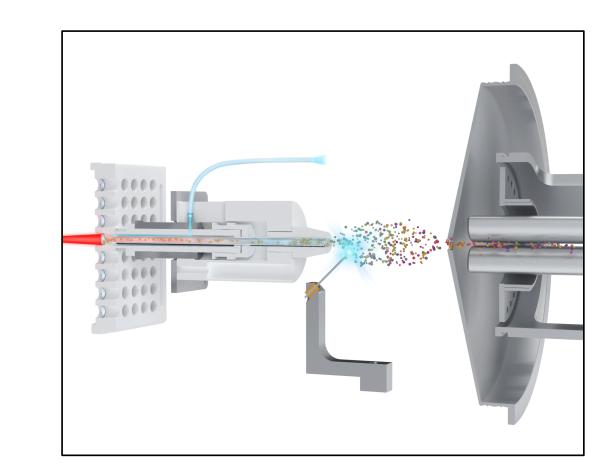


Figure 1 Schematic of the LDTD® ionization source

METHOD

1) Pre-Wash

- 10 mg Negative Hair sample
- 2 mL Dichloromethane
- Sonicate 5 min
- Remove Dichloromethane
- 2 mL Ethanol
- Sonicate 5 minutes
- Remove Ethanol

2) Hair Digest process

- 10 mg Hair sample (pre-washed)
- 100 µL Working solution (for standards)
- 100 µL Internal standard (5 ng/mL)
- 1 mL Methanol/TFA (9:1)
- Sonicate 1 hour
- React 8 hours at room temperature
- Transfer Organic phase in glass tube and evaporate to dryness
- 100 μL MeOH
- 900 μL HCL 1N/H₂O (1:14)
- Vortex
- SPE

Instrumentation

- LDTD® model S-960, Phytronix Technologies
- QTRAP® 5500 Systems, AB Sciex

LDTD Parameters

- Laser power pattern
- ➤ Increase laser power to 45 % in 6.0 s
- > Hold for 2 seconds
- ➤ Decrease laser power to 0 %
- Carrier gas flow: 3 L/min (Air)
- APCI (+) positive

LC Parameters

- ESI (+) positive
- Column: SilliaChrom SB-C18, 5µm (4.6x200)
- Flow rate: 0.5 ml/min
- MPA: Water/MeOH/FA (90/10/1)
- MPB: Water/MeOH/FA (10/90/1)
- Gradient: Time 30

MS Parameters

- Scan time: 10 msec
- DP: 100

3) SPE (Sample Cleanup, 100 mg/1cc)

Activation

- 1 mL EtAc/IPA/NH₄OH (80:15:5)
- 1 mL MeOH
- 1 mL Water

Load

 1 mL Sample Wash

- 1 mL Water
- 1 mL CH₃COONa (100 mM, pH 4.5)
- 1 mL IPA

Elution

Table 1 MRM transitions of drugs

Compound

6-Acetylmorphine (Quan)

6-Acetylmorphine (Conf)

Codeine (Quan)

Codeine (Conf)

Morphine (Quan)

Morphine (Conf)

Hydromorphone (Quan)

Hydromorphone (Conf)

Hydrocodone (Quan)

Hydrocodone (Conf)

Oxycodone (Quan)

Oxycodone (Conf)

Oxymorphone (Quan)

Oxymorphone (Conf)

Codeine-D6

Hydrocodone-D6

Hydromorphone-D6

Oxycodone-D6

6-Acetylmorphine-D6

Morphine-D6

Oxymorphone-D3

- 1.5 mL EtAC/IPA/NH₄OH (80:15:5)
- Evaporate to dryness
- 100 μL MeOH/H₂O (75:25)
- Spot 4 µL in LazWell™ plate

Q1

328

328

300

300

286

286

315

315

329

329

345

345

331

331

306

335

334

292

334

284

Analyze after complete solvent evaporation

Q3

201

CE (V)

Linearity results

For each drug, a calibration curve ranging from 5 to 100 ng/mL has been prepared in MeOH and spiked in negative hair samples. All curves are linear and show a good correlation coefficient, R²≥0.9967. Figure 2 shows the calibration curve results for 6-AM using LDTD® (R=0.99768). Table 2 shows the accuracy and precision results for 6-Acetylmorphine samples.

RESULTS

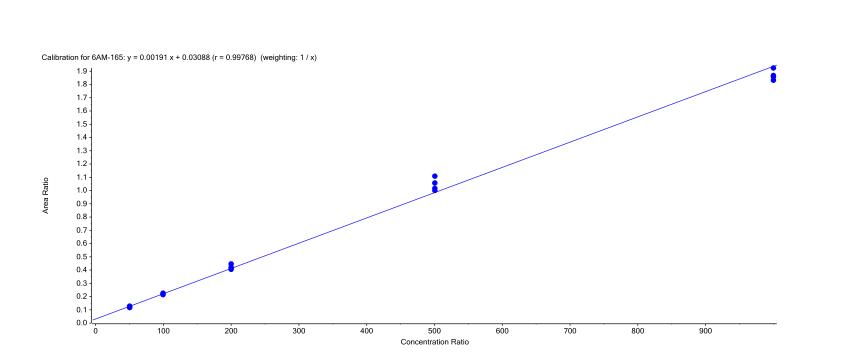


 Table 2 Precision and accuracy for 6-AM curve samples

Conc (ng/mL)	Conc (pg/mg)	Mean (pg/mg)	N	%RSD	%NOM
5	50	47.5	4	6.88	95.0
10	100	99.2	4	3.00	99.2
20	200	205.2	4	4.17	102.6
50	500	533.5	4	4.73	106.7
100	1000	964.6	4	2.16	96.5

Cross Validation LC vs LDTD®

Positive and negative real patient hair samples were analyzed both with LC-MS/MS and LDTD®-MS/MS to validate the efficiency of the LDTD® analysis method. No false positive results were obtained with the negative hair samples. Positive hair samples correlation results (pg/mg hair) are shown in Figure 3 to 8. No positive samples (higher the LC-MS/MS cut off) are available for OXM.

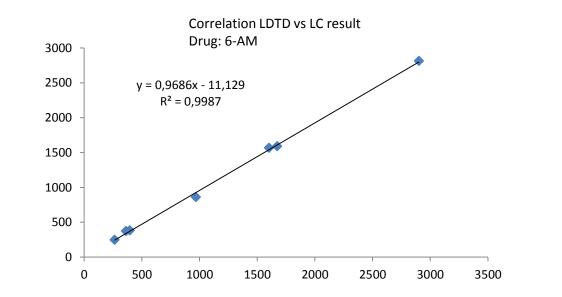


Figure 2 Calibration curve for 6-AM drug

Figure 3 LC vs LDTD® cross validation results for 6-AM

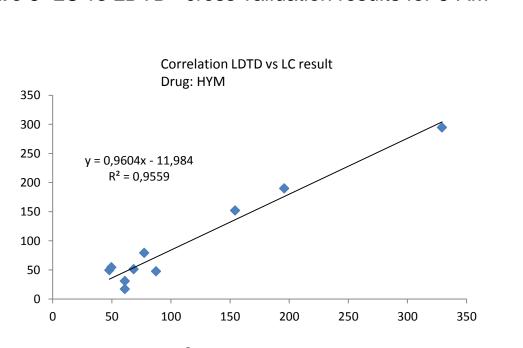


Figure 6 LC vs LDTD® cross validation results for HYM

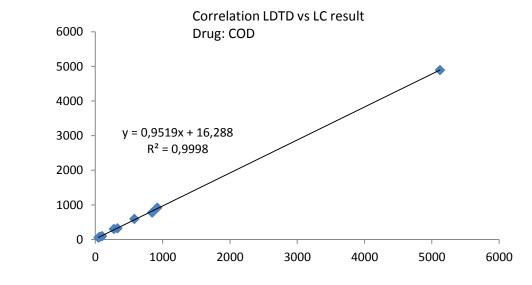
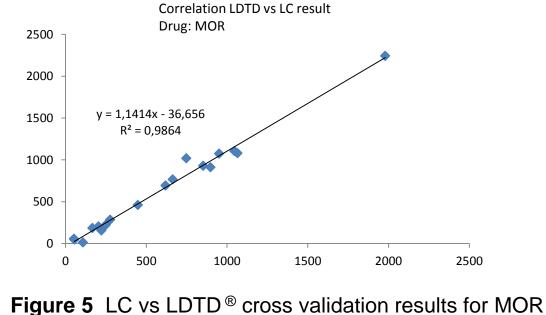


Figure 4 LC vs LDTD® cross validation results for COD

Drug: HYC

y = 0.8958x + 27.663 $R^2 = 0.9691$

Correlation LDTD vs LC result



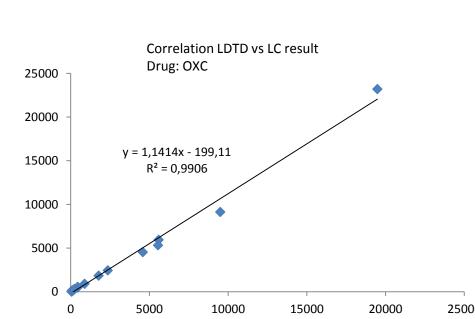


Figure 8 LC vs LDTD® cross validation results for OXC

CONCLUSIONS

Figure 7 LC vs LDTD® cross validation results for HYC

- LDTD® provides Ultra-Fast High-Throughput analysis of sample extract in 9 seconds sample-to-sample without any carry over.
- LC-MS/MS and LDTD®-MS/MS values agree for cross validation of real patient samples.
- A) Salary/Consultant Fees: Phytronix