

## INTRODUCTION

Testing for drugs of abuse in oral fluids can strongly benefit the criminal justice field as a less invasive and cost-effective approach for drug detection when compared to blood or urine sampling. Oral fluid analysis has facilitated laboratory analysis for many drugs of abuse and is a constantly evolving analysis procedure which benefits from increasingly sensitive methods of detection. The Laser Diode Thermal Desorption (LTD) source combined with Mass spectrometry is presented as a new screening tool for drug analysis in Oral Fluids. Analysis speed of LTD provides accurate results in seconds in combination with exceptional specificity of MS instruments make a powerful platform for the screening of different drugs of abuse and new emerging drugs. Different extraction procedures are available; however those methods depend on specific drug conditions: basic or acid drugs / hydrophilic or hydrophobic. A new extraction approach, Supported Liquid Extraction (SLE+) is evaluated as generic extraction procedure.

### LTD Ionization Source:

The LTD uses a Laser Diode to produce and control heat on the sample support which is a 96 well plate. The energy is then transferred through the sample holder to the dry sample which vaporizes prior to being carried by a gas in an APCI region. High efficiency protonation with strong resistance to ionic suppression characterize the ionization due to the absence of solvent and mobile phase. This allows very high throughput capabilities of 9 seconds sample-to-sample analysis time, without carry over.

## METHOD

### Sample preparation:

Supported Liquid Extraction (SLE):  
ISOLUTE-SLE+ (400µL)

- 200 µL sample
  - 10 µL of internal standard (IS)
  - 200 µL Ammonium hydroxide (1% in water)
  - Load onto SLE cartridge. A pulse of vacuum or positive pressure is applied to initiate flow and allow the sample to adsorb for 5 minutes.
  - Addition of 1 ml of ethyl acetate to elute. Wait 5 minutes.
  - Addition of 1 ml of dichloromethane/IPA (95/5) and allow to flow under gravity.
  - 6 µL of elution is directly deposited in the Lazwell plate.
- Dry samples prior to analysis

### Instruments setting:

Mass spectrometer AB Sciex 5500 QTrap operated in APCI positive mode.

MRM transitions used with DP=100, 3 µA discharge current and 5 msec dwell time

LTD model S-960 operated with a gas flow rate of 3 L/min and a laser pattern ramp from 0 to 45% in 3 seconds, maintaining this power level for 2 seconds before dropping it back to 0.



Figure 1: LTD system on AB SCIEX 5500 Qtrap® Mass Spectrometer

### Drug cut-off

Standard solutions are spiked in Orasure buffer at 0.5X, 1X and 2.5X the cut off level. The following cut-off levels are reported in Table 1.

Compound	Cut-Off
	(ng/ml)
Amphetamine	10
Butalbital	20
BZE	10
Cocaine	2
Cotinine	10
Fentanyl	10
MDA	10
MDEA	10
MDMA	10
Methamphetamine	10
Methadone	5
Phenobarbital	20
Secobarbital	20

Table 2 MRM transitions

Table 1 Screening cut-off

## RESULTS

### Intra-Run reproducibility and accuracy

Reproducibility and accuracy intra-run result at cut-off level are reported in Table 3.

Compound	Conc. (ng/ml)	N	Mean (ng/ml)	%RSD (%)	%Nom (%)
Amphetamine	10	6	10,6	18,5	106,4
Butalbital	20	6	20,0	9,2	99,9
BZE	10	6	11,0	9,0	109,9
Cocaine	2	6	2,0	4,4	100,8
Cotinine	10	6	9,7	3,2	97,4
Fentanyl	10	6	10,2	4,1	102,1
MDA	10	6	10,4	9,9	103,9
MDEA	10	6	10,3	4,7	103,2
MDMA	10	6	10,3	6,5	102,6
Methamphetamine	10	6	10,0	5,9	100,5
Methadone	5	6	5,1	7,3	101,8
Phenobarbital	20	6	19,1	3,4	95,3
Secobarbital	20	6	20,5	6,2	102,3

Table 3 Intra-run reproducibility and accuracy.

### Wet stability of extract sample

Following the extraction, sample are keep at 4°C. Samples were re-analyzed after 24 hours.

Reproducibility and accuracy result are reported in Table 5.

Compound	Conc. (ng/ml)	N	Mean (ng/ml)	%RSD (%)	%Nom (%)
Amphetamine	10	3	9,7	4,7	97,4
Butalbital	20	3	20,6	13,4	102,6
BZE	10	3	11,0	13,7	109,6
Cocaine	2	3	2,3	6,9	113,0
Cotinine	10	3	9,7	9,0	96,7
Fentanyl	10	3	11,4	1,9	114,4
MDA	10	3	10,7	7,4	107,2
MDEA	10	3	9,6	7,0	96,2
MDMA	10	3	10,0	11,2	100,1
Methamphetamine	10	3	9,6	9,5	96,3
Methadone	5	3	4,8	2,4	94,8
Phenobarbital	20	3	19,4	12,7	96,8
Secobarbital	20	3	19,6	8,6	98,5

Table 5 Wet stability result

### Linearity

Inter-run linearity results are reported in Table 8 and typical standard curve are presented in Figure 3.

Compound	Run 1	Run 2	Run 3
	r	r	r
Amphetamine	0,9966	0,9939	0,9950
Butalbital	0,9976	0,9923	0,9935
BZE	0,9959	0,9923	0,9944
Cocaine	0,9958	0,9969	0,9914
Cotinine	0,9975	0,9952	0,9962
Fentanyl	0,9956	0,9938	0,9970
MDA	0,9967	0,9973	0,9935
MDEA	0,9970	0,9983	0,9970
MDMA	0,9984	0,9957	0,9930
Methamphetamine	0,9985	0,9951	0,9923
Methadone	0,9977	0,9979	0,9934
Phenobarbital	0,9954	0,9908	0,9930
Secobarbital	0,9950	0,9926	0,9924

Table 8 Linearity result

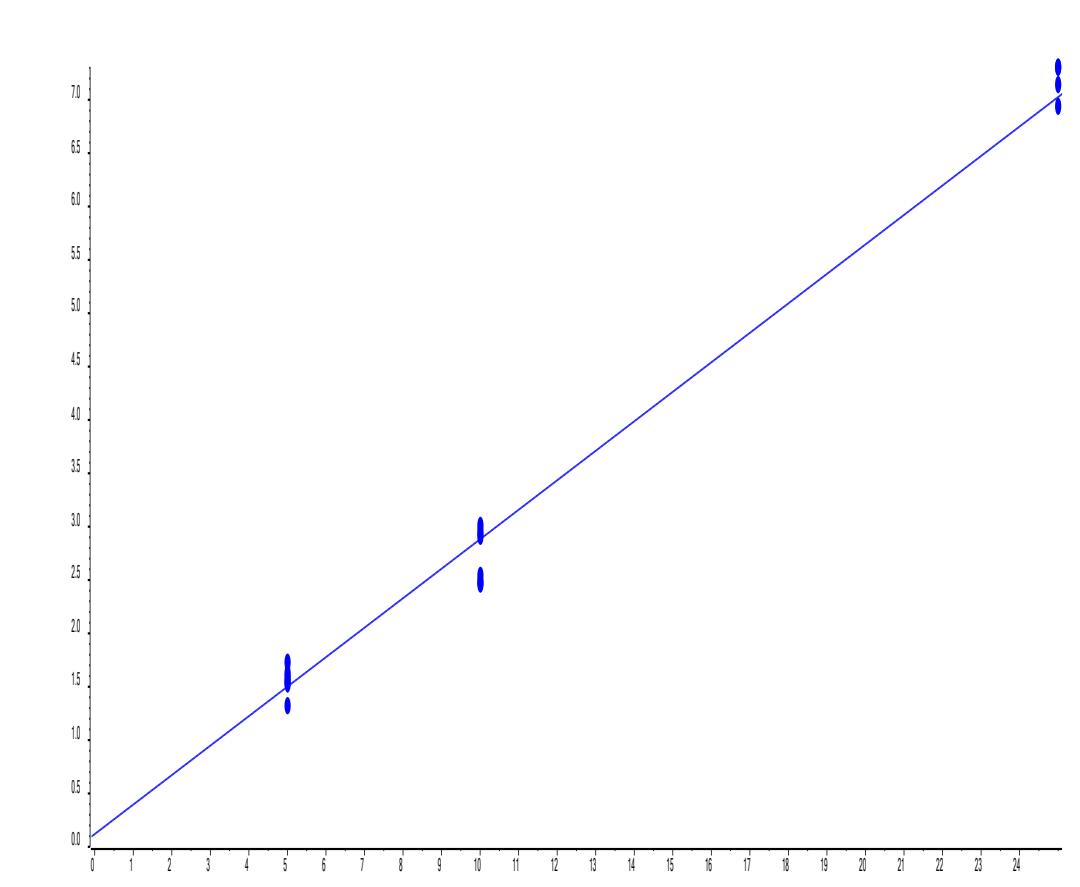


Figure 3 Typical standard curve

### Inter-Run reproducibility and accuracy

Reproducibility and accuracy inter-run result at cut-off level are reported in Table 4.

Compound	Conc. (ng/ml)	N	Mean (ng/ml)	%RSD (%)	%Nom (%)
Amphetamine	10	18	10,6	13,8	105,5
Butalbital	20	18	21,0	12,5	105,2
BZE	10	18	10,6	8,9	105,7
Cocaine	2	18	2,0	5,1	100,3
Cotinine	10	18	10,0	5,0	100,2
Fentanyl	10	18	10,2	4,5	101,9
MDA	10	18	10,1	7,8	100,8
MDEA	10	18	10,2	4,0	101,5
MDMA	10	18	10,2	5,5	101,5
Methamphetamine	10	18	10,1	5,2	101,5
Methadone	5	18	5,0	5,6	99,8
Phenobarbital	20	18	20,0	9,8	99,8
Secobarbital	20	18	21,0	11,0	105,2

Table 4 Inter-run reproducibility and accuracy.

### Recovery result

Recovery results are reported in Figure 2

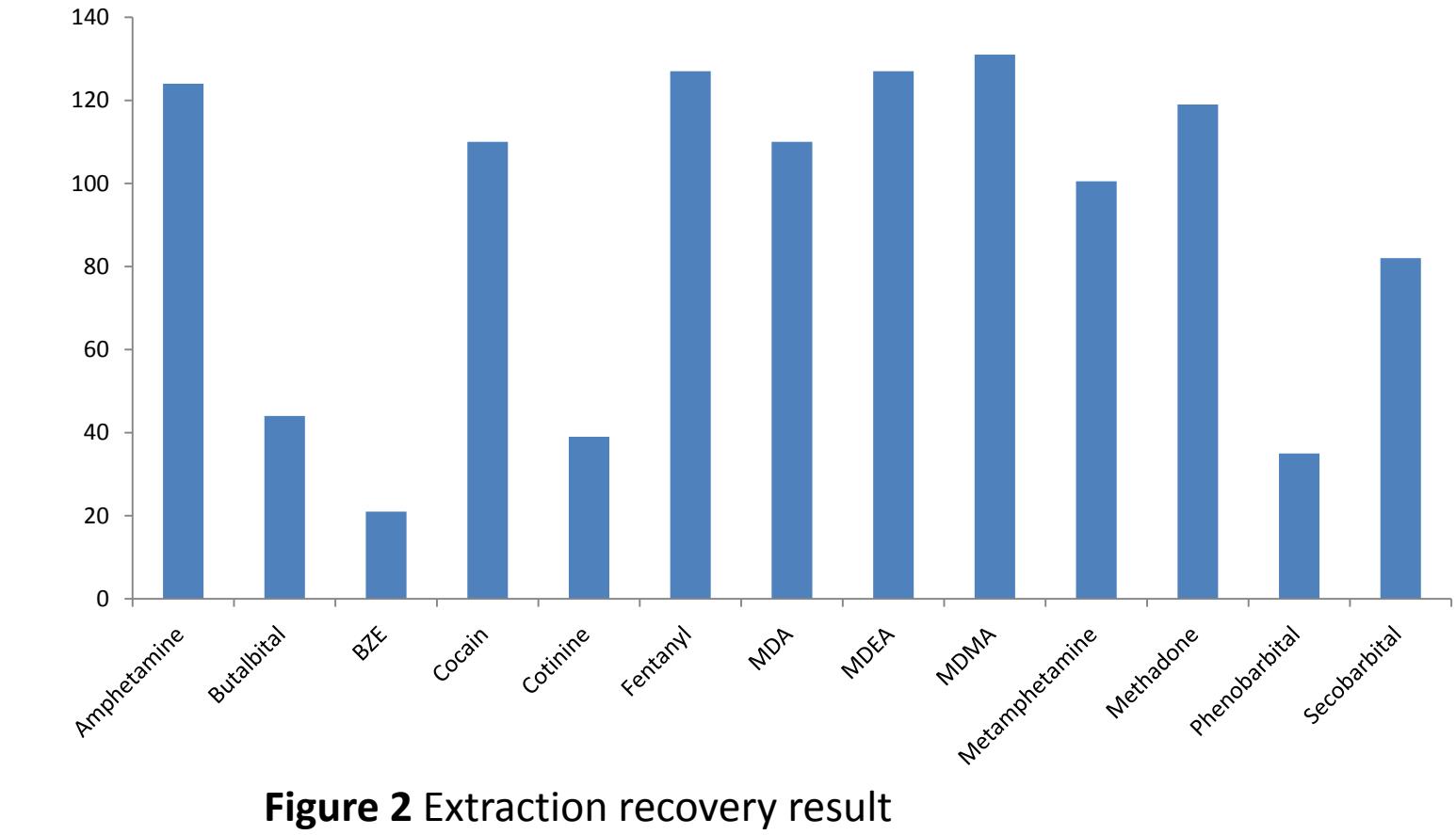


Figure 2 Extraction recovery result

### Common drug interferences

Common drug interferences are evaluated by adding a cocktail of known drugs to the QC .Results are reported in Table 7.

Compound	M1	M2	M3	M4	M5	M6
Amphetamine	Mean 11,80	11,20	9,20	9,10	8,80	8,50
	%RSD 12,5	3,3	7,4	15,2	14,0	7,8
	%Nom 118,0	112,0	92,0	91,0	88,0	85,0
Butalbital	Mean 18,90	20,70	21,30	22,05	20,10	24,90
	%RSD 9,9	14,2	10,6	14,9	3,6	5,7
	%Nom 94,5	103,5	106,5	110,3	100,5	124,5
BZE	Mean 9,00	9,20	9,60	8,80	9,90	10,80
	%R					